Pd(II)-Catalyzed Direct β-Arylation of C(sp³)-H Bonds of Aliphatic and Alicyclic Carboxamides *Via* **One-pot Method**

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Certificate of Examination

This is to certify that the dissertation titled "*Pd(II)-Catalyzed Direct β-Arylation of C(sp³)-H Bonds of Aliphatic and Alicyclic carboxamides Via One-pot Method*" submitted by Ms. Sruthi Mohan (Reg. No. MS11034) for the partial fulfillment of BS-MS dual degree programme of the Institute, has been examined by the thesis committee duly appointed by the Institute. The committee finds the work done by the candidate satisfactory and recommends that the report be accepted.

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Declaration

The work presented in this dissertation has been carried out by me under the guidance of **Dr. S. Arulananda Babu** at the Indian Institute of Science Education and Research Mohali.

This work has not been submitted in part or in full for a degree, a diploma, or a fellowship to any university or institute. Whenever contributions of others are involved, every effort is made to indicate this clearly, with due acknowledgement of collaborative research and discussion. This thesis is a bonafide record of original work done by me and all sources listed within have been detailed in the bibliography.

> Sruthi Mohan (Candidate)

Dated : April 22, 2016

In my capacity as the supervisor of the candidate's project work, I certify that the above statement by the candidate are true to the best of my knowledge.

> Dr. S. Arulananda Babu (Supervisor)

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Abstract

A step economical one-pot multicomponent reaction protocol comprising the installation of the bidentate directing group followed by the Pd(II)-catalyzed C-H activation and β-arylation of $C(sp^3)$ -H bonds of carboxamides is reported. Accordingly, the reaction of a mixture of an acid chloride, bidentate ligand (e.g., 8 aminoquinoline), and an aryl iodide in the presence of the $Pd(OAc)_2$ catalyst and Ag2CO³ additive directly afforded the corresponding β-C-H -arylated *N*-(quinolin-8 yl) carboxamide derivatives.

Chapter 1 Introduction

 The most demanding synthetic method in organic chemistry is the invention and improvement of carbon-carbon bond and carbon-heteroatom bond formation. Transition metal catalyzed synthetic methods, especially cross-coupling reactions^{1, 2} have made a great impact on organic chemistry. These methods revolutionized the field of organic chemistry for discovering novel reactions with yield and efficiency. General reaction mechanism (Scheme 1) of cross-coupling reactions starts with the oxidative addition of an electrophile **2** to the metal **1** generates the intermediate **3**. Next, transmetallation of organometallic reagent **4** with **5** gives the intermediate **6**. Last step reductive elimination of intermediate **6** gives the C-C coupled product **7**. Cross-coupling reactions have some limitations such as a) it requires pre-functionalized starting materials, b) use of costly and bulky ligands, c) production of hazardous side products. Therefore, C-C bond and C-X bond formation *via* C-H activation methodology is considered as the most demanding synthetic strategy.

 Carbon-Hydrogen (C-H) activation is the process of activation of a particular C-H bond using a low valent transition metal in the presence of oxidants or additives³. C-H functionalization is the reaction in which activation of C-H bond, cleavage and replacement of C-H bond to C-X bond⁴ (X can be carbon, nitrogen, oxygen) (figure 1). Transition metal-catalyzed C-H activation followed by the C-C

coupling reaction is one of the remarkable synthetic transformations in organic synthesis 5 .

 Carbon-Hydrogen bond is ubiquitous, often inert and largely found in organic molecules. C-H bond have high dissociation energy 85-120 Kcal/mol and non-polarizable in nature. Utilization of this C-H bond was a challenging task in earlier times. So C-H activation methodology gained much importance in the field of organic chemistry. C-H activation has been well explored during last decades because of its advantages like a) do not require pre-functionalized starting material and b) do not generate any hazardous by-products. Many complex molecules can be easily synthesized from simple starting material through activating the C-H bond (Scheme 2). C-H activation is the complementary tool for the total synthesis of complex natural products⁶.

Different Methods of C-H Functionalization

 Two different methods of C-H functionalizations are known. a) Direct functionalization of inert C-H bonds and b) directing group assisted C-H functionalization. In first method, there is no regioselectivity, because all the C-H bonds have the equal possibility to undergo C-H activation^{7, 8}. But in directing group assisted C-H activation, the metal center will coordinate with the heteroatom thereby activates the proximal C-H bond leaving other C-H unreactive \degree . (Scheme 3)

 Three classes of directing groups (DG) are reported till now. They are a) non-modifiable DG, b) modifiable or transformable DG and c) reusable $DG⁹$. Nonmodifiable DG cannot be modified in to any other functionality¹⁰. But modifiable DGs can be transformed in to desired functionality after the completion of reaction 11 . Pre-installed chelating groups in to the starting material can be readily recovered from the C-H functionalized product are termed as reusable DGs. In this project we explored the use of reusable DGs^9 .

One of the major challenges in this field is the installation of DGs plays a key role in C-H activation processes. Present emergency is to develop a complementary technology to enhance the efficiency of DG-based C-H activation transformations. In this regard, *Wan et al*. reported a multicomponent reaction method of Pd-catalyzed C-H arylation of sp^2 C-H bonds of aromatic carboxamides¹². Understanding the importance of the bidentate ligand directed Pd-catalyzed $sp³$ C-H functionalization of carboxamides in synthetic organic chemistry, we developed a simple methodology to enhance the efficiency of Pd-catalyzed $sp³$ C-H functionalization reactions^{13,} ¹⁴. We report a one-pot multicomponent reaction protocol comprising installation of

bidentate DG, Pd-catalyzed C-H activation and β -arylation of C(sp³)-H bonds of various aliphatic and alicyclic carboxamides (scheme 4).

Cyclobutane is the second smallest, strained four-membered carboxylic ring. It presents as the core unit of many natural products (Figure 2), synthetic compounds and many pharmaceutical agents^{15, 16}. Monoarylated and bis-arylated cyclobutane carboxamides exhibit a wide range of biological activities and medicinal properties¹⁷. This type of compounds can be easily synthesized by one-pot multicomponent reaction methodology of Pd-catalyzed C-H activation of $sp³$ bonds of alicyclic carboxamides. Substituted fatty acids can also be easily synthesized *via* Pd-catalyzed C-H activation on $sp³$ bonds of aliphatic carboxamides.

Chapter 2 Results and Discussions

 We carried out optimization studies to get the best reaction conditions for establishing a one-pot reaction protocol comprising the installation of bidentate ligand and β-arylation on sp^3 C-H on cyclobutane based carboxamides. Multicomponent reaction of a mixture of bidentate ligand 8-aminoquinoline (**9a**), cyclobutanecarbonyl chloride $(8a)$ and PhI $(11a)$, Pd (OAc) ₂ catalyst and Ag salt as an additive represents the Table 1. We carried out the reaction of a mixture of 8 aminoquinoline (**9a**), cyclobutanecarbonyl chloride (**8a**) and PhI (**11a,** 4 equiv), Pd(OAc)₂ catalyst and AgOAc additive in toluene at 110 ^OC, which resulted in the formation of bis-arylated cyclobutane carboxamide in 20% yield (Table 1, entry 1). To the same reaction, instead of toluene solvent *tert*-butanol was added .This reaction also afforded the bis-arylated cyclobutane carboxamide in only 20% yield (Table 1, entry 2). Next, we performed another reaction of a mixture of 8-aminoquinoline (**9a**), cyclobutanecarbonyl chloride (8a) and PhI (11a, 8 equiv), Pd(OAc)₂ catalyst, AgOAc and K_2CO_3 as additive in *o*-xylene at 120^oC failed to give the product **12a** (Table 1, entry 3). Again, we carried out the same reaction with 4 equiv of PhI, instead of AgOAc additive Ag_2CO_3 was used. Here we observed the formation of bis-arylated product in comparatively good yield ie, 59% (Table 1, entry 4).

After observing the use of Ag_2CO_3 instead of AgOAc improved the yield, next we performed some reactions using Ag_2CO_3 as additive. Multicomponent reaction of a mixture of 8-aminoquinoline (**9a**), cyclobutanecarbonyl chloride (**8a**) and PhI (11a, 4 equiv), Pd(OAc)₂ catalyst and Ag₂CO₃ additive in *t*-amylOH at 80^oC also afforded the same bis-arylated product in 68% yield (Table 1, entry 5). The same reaction in *o*-xylene instead of *t*-amylalcohol also resulted in the same product with yield 62%.(Table 1, entry 6). Then tried the same reaction by increasing the equivalent of PhI to 8 equiv in o-xylene solvent and gave the product of 82% yield (Table 1, entry 7). Then we tried one reaction with K_2CO_3 additive instead of Ag_2CO_3 , which failed to afford the product **12a** (Table 1, entry 8).

^a The reaction period for all the reactions is 36 h. $\frac{b}{c}$ The reaction period is 12 h. $\frac{c}{c}$ The reaction period is 24 h.

Then the reaction of **9a**, **8a**, PhI (11a, 4 or 6 equiv) and Ag_2CO_3 additive in the presence of 5 mol% of the $Pd(OAc)_2$ catalyst instead of 10mol% catalyst gave the product **12a** in 54-75% yields (Table 1, entries 9 and 10). In another trial, the mixture of **9a**, **8a** and PhI (11a, 8 equiv), Pd(OAc)₂ catalyst in the presence of Ag2CO3 and Cs2CO3 additives gave the product **12a** in only 28% yield (Table 1, entry 11). In similar trial, instead of the mixture of additives we used the Ag_2CO_3 under neat condition resulted in the formation of product **12a** in a improved yield 58% (Table 1, entry 12). The same reaction in *tert*-amylOH instead of *o*-xylene also afforded the product **12a** in 66-72% yield (Table 1, entries 13 and 14).

From these optimization reactions, it is clear that the one-pot Pd-catalyzed C-H arylation reaction of a mixture of 8-aminoquinoline (**9a**), cyclobutanecarbonyl chloride (8a) and PhI (11a, 8 equiv), Pd(OAc)₂ catalyst successfully afforded the product $12a$ only in the presence of Ag_2CO_3 additive.

 In the conventional reaction condition for the synthesis of bis-arylated cyclobutane carboxamide derivatives AgOAc was used as the additive, which helps to regenerate the Pd(OAc)₂ catalyst in the proposed Pd^H-Pd^{IV} catalytic cycle. In this multicomponent reaction methodology, AgOAc is not favouring for C-H arylated product $12a$ in satisfactory yield. But Ag_2CO_3 is promoting the DG installation reaction as well as C-H arylation reaction.

 After the optimization of reaction conditions, we investigated the scope and generality of this multicomponent reaction in one-pot method to synthesize different varieties of bis-arylated cycloalkane carboxamide derivatives. We carried out some reactions of Ag_2CO_3 mediated, Pd-catalyzed C-H activation protocol involving 8aminoquinoline (**9a**), cyclobutanecarbonyl chloride (**8a**) and a variety of aryl iodides,

which resulted in the formation of bis-arylated cyclobutane carboxamide derivatives **12a-l** (Table 2). We performed certain reactions with aryl iodides containing electron withdrawing and donating groups at the *para* position afforded the product **12b-g** in low to high yields (40-83%, Table 2). Reaction with 1-iodo-4-nitrobenzene gave the product **12b** in low yield (40%, Table 2) may be due to the low reactivity which contains a strong electron-withdrawing group (nitro group) at the *para* position. Same reaction with aryl iodide containing the CF_3 group at meta position gave the product **12g** in 80% yield (Table 2).

 Multicomponent reactions with two different aryl iodides like 1 iodonaphthalene and 6-iodo-2,3-dihydrobenzo[*b*][1,4] dioxine gave the corresponding products **12h** (25%) and **12i** (10%) in only low yields. Low yields in these reactions may be due to the low reactivity of the corresponding aryl iodides. Next, we performed Ag_2CO_3 mediated, Pd-catalyzed C-H activation reactions involving 8aminoquinoline (**9a**), cyclobutanecarbonyl chloride (**8a**) and di-substituted aryl iodides afforded the corresponding products **12j** (71%) and **12k** (60%) (Table 2). C-H arylation protocol using the heteroaryl iodide such as 2-iodothiophene with substrates 8-aminoquinoline (**9a**), cyclobutanecarbonyl chloride (**8a**) furnished the product **12l** in 78% yield (Table 2).

 We also investigated the one-pot C-H arylation reactions on different substrates such as cyclohexanecarbonyl chloride and cyclopropanecarbonyl chloride. In the same Pd-catalyzed, Ag_2CO_3 mediated C-H arylation reaction, instead of cyclobutanecarbonyl chloride (**8a**), cyclohexanecarbonyl chloride was used along with 8-aminoquinoline (**9a**) and different aryl iodides like PhI (**11a**) and 4-iodoanisole furnished the products **12m** (38%), **12n** (35%) (Table 2). Traces of by-products are also obtained that we could not isolate in pure form. Low yield in cyclohexane system may be due to the increasing steric crowding and the C-H arylation seems to be sluggish here. We performed the Pd-catalyzed, Ag_2CO_3 mediated C-H arylation reaction in cyclopropane system furnished the product **12o** (40%) (Table 2). This low yield may be because the C-H arylation of cyclopropane reaction seems to be sluggish in nature.

 Successively, we elaborated the scope and generality of this method by using various aliphatic carbonyl chlorides. First we carried out some reactions of Pdcatalyzed, Ag_2CO_3 mediated C-H arylation protocol involving the butyryl chloride (**13a**), 8-aminoquinoline (**9a**) and aryl iodides containing various substituents at the *meta/para* position or disubstituted aryl iodides, which resulted in the formation of mono-arylated butane carboxamides **14a-o** in good to high yields (50-99%, Table 3). Further we elaborated this reaction methodology to check the substrate scope using different aliphatic carbonyl chlorides. Accordingly, various β -methylene C(sp³)-H arylated aliphatic carboxamides **14p-u** were obtained in high yields (80-97%, Table 4). We performed the same Pd-catalyzed, Ag_2CO_3 mediated C-H arylation protocol using propionyl chloride gave the corresponding mono arylated (**14v**) and bis arylated propanecarboxamide (**14v'**) derivatives in 17 and 58% yields (Table 4).

Next we carried out some reactions to find out the feasibility of using other bidentate ligands such as, 2-(methylthio)aniline (9b) and N^1 , N^1 -dimethylethane-1,2-diamine (9c) for this one-pot multicomponent reaction protocol comprising installation of bidentate DG followed by Pd-catalyzed C-H activation and arylation of $C(sp^3)$ -H bonds of carboxamides. One-pot Pd-catalyzed, Ag_2CO_3 mediated C-H arylation protocol involving bidentate ligands 2-(methylthio)aniline (9b) or N^1 , N^1 -dimethylethane-1,2diamine (**9c**), cyclobutanecarbonyl chloride (**8a**) and 4-iodoanisole failed to afford the bis-arylated cyclobutanecarboxamides **12p** and **12q** (Scheme 5). Based on the reactions in Table 1, 2, 3 and 4, the bidentate ligand 8-aminoquinoline (**9a**) found to be the best for accomplishing the one-pot multicomponent reaction protocol comprising installation of bidendate DG followed by Pd-catalyzed C-H activation and arylation of $C(sp^3)$ -H bonds of carboxamides.

 We have also used the condition reported by Quin *et al*. Pd-catalyzed, PhI(OAc)² mediated C-H arylation protocol involving 8-aminoquinoline (**9a**), cyclobutanecarbonyl chloride (**8a**) and PhI (**11a,** 2.5 equiv) also afforded the product **12a** in 40% yield (Scheme 5).

We noted that the reagents for alkylation and arylation are different in literature reports and the arylation of sp^3 C-H bonds are more easier than alkylation of sp^2 C-H bonds of carboxamides. We tried one alkylation reaction on $sp³$ C-H bond using the reported condition involving 8-aminoquinoline (**9a**), cyclobutanecarbonyl chloride (**8a**) and *n*-butyl iodide failed to afford the C-H alkylated cyclobutanecarboxamide **12r** (Scheme 5). But we got the corresponding product **12s (**15-25%) of alkylation reaction in another trial using the condition reported by Daugulis group on propionyl based system. It should be noted that the alkylation condition needs a base, such as $K₂CO₃$ and the low yield may be due to the side reaction of N-alkylation of bidentate ligand. In order to examine whether the alkylation of C-H bonds will work or not, we tried one reaction involving 8-aminoquinoline (**9a**), benzoyl chloride and *n*-butyl iodide or *n*-heptyl iodide. Notably, this multicomponent-based $sp²$ alkylation reactions were successful and the products **12t** (45%) and **12u** (60%) were obtained in satisfactory yield.

Removal of bidentate DGs from C-H arylated carboxamides has also been successfully achieved. In the conventional method, the starting material C-H arylated carboxamides usually isolates through column chromatography. Here we attempted to avoid the column chromatographic purification step of crude reaction mixture and directly performed the hydrolysis reaction of amide bond from the crude reaction mixture. The NaOH mediated amide hydrolysis or BF_3 . OEt₂ mediated esterification of the carboxamides resulted in the formation of products **15a** (40%), **15b** (32%), **15c** (52%), and **15d** (59%) in satisfactory yields (Scheme 6).

Chapter 3

Experimental Methods

General. Melting points of compounds are uncorrected. ${}^{1}H$ and ${}^{13}C$ NMR spectra were recorded on 400 MHz and 100 MHz spectrometers respectively with TMS as an internal or external standard. IR spectra were recorded as KBr pellets or thin films. Column chromatographic purification of the crude reaction mixtures was performed using silica gel (100-200 mesh). HRMS measurements of compounds reported in this work were obtained from QTOF mass analyzer using electrospray ionization (ESI) technique. Reactions were carried out in dry solvent under nitrogen atmosphere wherever required. Solutions were dried using anhydrous sodium sulfate. Reagents were added to the reaction flask with the help of syringe. Thin layer chromatography (TLC) was performed on silica plates or and components were visualized by observation under iodine vapor. Isolated yields of all the products were reported (yields were not optimized). Characterization of the known compounds was carried out by comparing the data with the literature data.

General Procedure A for the synthesis of carboxamides

8-Aminoquinoline (0.4 mmol) and aryl iodide (4-6 equiv.) were taken in a 10 mL round bottom (RB) flask attached with a condenser under N_2 atmosphere. To this mixture, Pd(OAc)₂ catalyst (10 mol%), Ag₂CO₃ additive (1.5-2 equiv.) and cyclobutanecarbonyl chloride (0.4 mmol) were added. Finally, added the solvent oxylene (2 mL) in to the RB and sealed the condenser with a stopper. The mixture was stirred at 120 $\rm{^{\circ}C}$ for 36 h. After the completion of reaction, the mixture was allowed to cool in room temperature. The solvent was removed at reduced pressure. The residue was subjected to silica gel column chromatography to give pure products by using hexane and ethyl acetate (5:0.25).

Procedure B: Hydrolysis of amide bond using esterification method.

Procedure A was used to synthesize the carboxamide. After the completion of reaction, removal of solvent was done under reduced pressure and hydrolysis procedure was done on dried crude reaction mixture. Column chromatography purification step is not required. To the dried reaction mixture, MeOH (2 mL) was added and stirred vigorously under nitrogen atmosphere for 5 min. Finally, BF_3OEt_2 (1 mL) was added in to the reaction mixture and allowed to stir for 24 h at 85 $^{\circ}$ C. After the completion of reaction, the mixture is quenched by Triethyl amine and removed the solvent under reduced pressure. The residue was subjected to silica gel column chromatography to give pure products by using hexane and ethyl acetate $(5:0.1)$.

Procedure C: Hydrolysis of amide bond to corresponding acids

Procedure A was used to synthesize the carboxamide. After the completion of reaction, removal of solvent was done under reduced pressure and hydrolysis procedure was done on dried crude reaction mixture. Column chromatography purification step is not required. To the dried reaction mixture, ethanol (3 mL) was added and stirred vigorously under open atmosphere. NaOH (13 mmol) was added to the reaction mixture and was allowed to stir it for 24 h at 85 $\mathrm{^{0}C}$. After the reaction, transferred the reaction mixture in to a separating funnel and extracted with diethyl ether. Water layer was collected separately and that was basic in nature. Con.HCL was added into the aqueous layer to make it acid nature. Then again extracted with diethyl ether and collected the organic layer separately. The combined organic layers were dried over anhydrous Na₂SO₄. Thus the desired carboxylic acid was successfully obtained after the removal of solvent in high vacuum.

2,4-diphenyl-N-(quinolin-8-yl)cyclobutanecarboxamide (12a): Following the general procedure A described above, **12a** was obtained after purification by silica gel column chromatography (EtOAc : Hexane $= 4 : 96$) as brown color solid (77 mg, 82%); R_f 0.40 (15% EtOAc/Hexane); mp 145-147 ^oC; IR (DCM): υ_{max} 3342, 1669, 1521, 1484, 993 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.54 (s, 1H), 8.75 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.6$ Hz, 1H), 8.31 (dd, $J_1 = 7.6$ Hz, $J_2 = 1.6$ Hz, 1H), 8.06 (dd, $J_1 = 8.0$ Hz, J_2 $= 1.6$ Hz, 1H), 7.41-7.26 (m, 8H), 7.25-7.22 (m, 4H), 7.12-7.09 (m, 2H), 4.19 (dd, J_I) $= 8.4$ Hz, $J_2 = 3.2$ Hz, 1H), 4.14-4.09 (m, 2H), 3.62-3.54 (m, 1H), 2.80-2.77 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 168.9, 147.8, 140.6, 138.2, 136.2, 134.2, 128.1, 127.7, 127.5, 127.0, 126.1, 121.3, 120.9, 116.4, 54.6, 38.1, 29.9; HRMS (ESI): *m/z* $[M + H]^{+}$ calcd for $C_{26}H_{23}N_{2}O$: 379.1810; found 379.1824.

2,4-bis(4-nitrophenyl)-N-(quinolin-8-yl)cyclobutanecarboxamide (12b): Following the general procedure A described above, **12b** was obtained after purification by silica gel column chromatography (EtOAc : Hexane $= 20$: 80) as brown color solid (46 mg, 40%); R_f 0.40 (30% EtOAc/Hexane); mp 182-184 ^oC; IR (DCM): υ_{max} 3344, 1682, 1596, 1391, 840 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): ¹H NMR (400 MHz, CDCl₃): δ 9.64 (s, 1H), 8.78-8.76 (m, 1H), 8.21-8.19 (m, 1H), 8.14-8.11 (m, 1H), 8.10-8.07 (m, 4H), 7.49-7.41 (m, 6H), 7.32 (d, *J* = 8.0 Hz, 1H), 4.32 (dd, $J_1 = 8.4$ Hz, $J_2 = 3.2$ Hz, 1H), 4.22-4.15 (m, 2H), 3.65-3.56 (m, 1H), 2.90-2.87 (m, 1H); ¹³C NMR (100 MHz, CDCl3): δ 167.7, 148.1, 148.0, 146.5, 138.0, 136.5, 133.3, 127.8, 127.6, 127.2, 123.5, 121.9, 121.7, 116.5, 54.5, 38.7, 30.0; HRMS (ESI): m/z [M + H]⁺ calcd for C₂₆H₂₁N₄O₅: 469.1511; found 469.1500.

2,4-bis(4-chlorophenyl)-N-(quinolin-8-yl)cyclobutanecarboxamide (12c): Following the general procedure A described above, **12c** was obtained after purification by silica gel column chromatography (EtOAc : Hexane $= 4 : 96$) as yellow colour solid (93 mg, 83%); R_f 0.40 (15% EtOAc/Hexane); mp 171-174 ^oC; IR (DCM): v_{max} 2988, 1698, 1598, 1351, 778 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.55 (s, 1H), 8.75 (d, *J* = 3.6 Hz, 1H), 8.32 (d, *J* = 7.6 Hz, 1H), 8.10 (d, *J* = 8.0 Hz, 1H), 7.44-7.39 (m, 2H), 7.34 (t, *J* = 8.0 Hz, 1H), 7.28 (d, *J* = 8.4 Hz, 4H), 7.19 (d, *J* = 8.0

Hz, 4H), 4.17-4.12 (m, 1H), 4.05-3.99 (m, 2H), 3.47 (q, *J* = 10.8 Hz, 1H), 2.77-2.69 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 168.5, 147.9, 138.9, 138.2, 136.3, 133.9, 131.9, 128.4, 128.2, 127.8, 127.3, 121.5, 121.4, 116.5, 54.4, 38.4, 30.1; HRMS (ESI): m/z [M + H]⁺ calcd for C₂₆H₂₁Cl₂N₂O: 447.1030; found 447.1010.

2,4-bis(4-acetylphenyl)-N-(quinolin-8-yl)cyclobutanecarboxamide (12d): Following the general procedure A described above, **12d** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $30:70$) as yellow color solid (89 mg, 78%); R_f 0.1 (30% EtOAc/Hexane); mp181-183 ^oC; IR (DCM): v_{max} 3345, 1605, 1524, 1391, 793 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.58 $($ s, 1H $)$, 8.76 (dd, J_I = 4.4 Hz, $J₂$ = 1.2 Hz, 1H $)$, 8.24 (d, $J = 7.6$ Hz, 1 Hz $)$, 8.09 (dd, J_I = 8.0 Hz, *J2* = 1.2 Hz, 1H), 7.81 (d, *J* = 8.0 Hz, 4H), 7.44-7.37 (m, 6H), 7.31-7.27 (m, 1H), 4.30-4.25 (m, 1H), 4.17-4.10 (m, 2H), 3.63-3.55 (m, 1H), 2.86-2.79 (m, 1H), 2.49 (s, 6H); ¹³C NMR (100 MHz, CDCl3): δ 197.9, 168.3, 148.0, 146.3, 138.1, 136.3, 135.1, 133.8, 128.3, 127.7, 127.2, 127.0, 121.5, 121.4, 116.4, 54.5, 39.0, 29.8, 26.5; HRMS (ESI): m/z [M + H]⁺ calcd for C₃₀H₂₇N₂O₃: 463.2021; found 463.2012.

2,4-bis(4-methoxyphenyl)-N-(quinolin-8-yl)cyclobutanecarboxamide (12e): Following the general procedure A described above, **12e** was obtained after purification by silica gel column chromatography (EtOAc : Hexane $= 4 : 96$) as yellow colour solid (88 mg, 80%); R_f 0.40 (15% EtOAc/Hexane); mp 146-149 ^oC; IR (DCM): v_{max} 2936, 1689, 1612, 1519, 825 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.49 (s, 1H), 8.73 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.6$ Hz, 1H), 8.37 (dd, $J_1 = 7.2$ Hz, $J_2 = 1.6$ Hz, 1H), 8.06 (dd, *J*¹ = 8.4 Hz, *J*² = 1.6 Hz, 1H), 7.40-7.29 (m, 7H), 6.77 (d, *J* = 8.4 Hz, 4H), 4.09-4.07 (m, 1H), 4.02-4.00 (m, 2H), 3.70 (s, 6H), 3.53-3.45 (m, 1H), 2.72-2.66 (m, 1H); ¹³C NMR (100 MHz, CDCl3): δ 169.2, 157.9, 147.8, 138.2, 136.1, 134.3, 132.7, 128.2, 127.7, 127.2, 121.3, 120.9, 116.4, 113.5, 55.1, 54.8, 38.5, 30.5; HRMS (ESI): m/z [M + H]⁺ calcd for C₂₈H₂₇N₂O₃: 439.2021; found 439.2019.

N-(quinolin-8-yl)-2,4-di-p-tolylcyclobutanecarboxamide (12f): Following the general procedure A described above, **12f** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $4:96$) as pale yellow color solid (71 mg, 71%); R_f 0.40 (15% EtOAc/Hexane); mp 156-160 °C; IR (DCM): υ_{max} 3359, 1689, 1595, 1484, 791 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.50 (s, 1H), 8.75-8.73 (m, 1H), 8.37 (dd, $J_1 = 7.2$ Hz, $J_2 = 1.6$ Hz, 1H), 8.07 (dd, $J_1 = 8.0$ Hz, $J_2 = 1.6$ Hz, 1H), 7.41-7.29 (m, 3H), 7.25 (d, *J* = 8.0 Hz, 4H), 7.03 (d, *J* = 7.6 Hz, 4H) 4.16-4.11 (m, 1H), 4.07-4.00 (m, 2H), 3.55-3.47 (m, 1H), 2.74-2.68 (m, 1H), 2.22 (s, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 169.1, 147.7, 138.2, 137.6, 136.1, 135.4, 134.3, 128.8, 127.7, 127.3, 126.9, 121.3, 120.8, 116.5, 54.6, 38.9, 30.2, 21.0; HRMS (ESI): *m/z* [M + H]⁺ calcd for $C_{28}H_{27}N_2O: 407.2123$; found 407.2117.

N-(quinolin-8-yl)-2,4-bis(3-(trifluoromethyl)phenyl)cyclobutanecarboxamide

(12g): Following the general procedure A described above, **12g** was obtained after purification by silica gel column chromatography ($EtOAc$: Hexane = 4:96) as brown color solid (101 mg, 80%); R_f 0.40 (15% EtOAc/Hexane); mp 114-116 ^oC; IR (DCM): v_{max} 3445, 1519, 1321, 1313, 989 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.56

(s, 1H), 8.75 (d, *J* = 4.0 Hz, 1H), 8.24 (d, *J* = 7.6 Hz, 1H), 8.08 (d, *J* = 8.4 Hz, 1H), 7.60-7.53 (m, 4H), 7.43-7.29 (m, 7H), 4.25-4.22 (m, 1H), 4.17-4.10 (m, 2H), 3.58 (dd, *J1* = 22.0 Hz, *J2* = 11.2 Hz, 1H), 2.84 (dd, *J1* = 10.0 Hz, *J2* = 2.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl3): δ 168.2, 148.0, 141.3, 138.1, 136.2, 133.7, 130.3 (q, *J* = 32 Hz) , 128.5, 127.7, 127.1, 126.8 (q, *J* = 271 Hz), 123.7 (q, *J* = 4 Hz), 123.1 (q, *J* = 4 Hz), 121.4, 121.4, 116.4, 54.2, 38.7, 29.9; HRMS (ESI): *m/z* [M + H]⁺ calcd for $C_{28}H_{21}F_6N_2O: 515.1558$; found 515.1536.

2,4-di(naphthalen-1-yl)-N-(quinolin-8-yl)cyclobutanecarboxamide (12h): Following the general procedure A described above, **12h** was obtained after purification by silica gel column chromatography ($EtOAc$: Hexane = 4:96) as brown color solid (30 mg, 25%); R_f 0.40 (15% EtOAc/Hexane); mp 225-227 ^oC; IR (DCM): v_{max}): 3351, 1683, 1523, 1485, 780 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.03 (br s, 1H), 8.42 (dd, 1H, *J*¹ = 4.2 Hz, *J*² = 1.6 Hz), 8.23 (d, 2H, *J* = 8.4 Hz), 7.84 (dd, 1H, *J*¹ $= 7.6$ Hz, $J_2 = 1.3$ Hz), 7.76 (dd, 1H, $J_1 = 8.4$ Hz, $J_2 = 1.5$ Hz), 7.69 (d, 2H, $J = 7.0$ Hz), 7.65 (d, 2H, *J* = 8.0 Hz), 7.58-7.51 (m, 4H), 7.42 (t, 2H, *J* = 8.0 Hz), 7.33 (t, 2H, *J* = 7.0 Hz), 7.16 (dd, 1H, *J*¹ = 8.2 Hz, *J*² = 4.2 Hz), 7.03-6.94 (m, 2H), 4.78-4.72 (m, 3H), 4.11-4.03 (m, 1H), 2.91-2.88 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 168.2, 147.3, 137.7, 135.8, 135.7, 133.7, 133.5, 131.8, 128.8, 127.2, 127.0, 126.7, 126.0, 125.4, 125.2, 125.0, 123.5, 120.9, 120.5, 115.8, 56.1, 38.0, 28.1;; HRMS (ESI): *m/z* $[M + H]^{+}$ calcd for $C_{34}H_{27}N_{2}O$: 479.2123; found 479.2115.

2,4-bis(2,3-dihydrobenzo[b][1,4]dioxin-6-yl)-N-(quinolin-8 yl)cyclobutanecarboxamide (12i): Following the general procedure A described

above, **12i** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 5 : 95) as brown semi solid (13 mg, 10%); R*^f* 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3333, 2935, 1681, 1510, 1485, 1384, 1162, 1068, 825 cm-1 ; ¹H NMR (400 MHz, CDCl3): *δ* 9.75 (s, 1H), 8.82 (dd, *J1* = 7.2 Hz, *J2* = 1.6 Hz, 1H), 8.71 (dd, *J1* = 4.0 Hz, *J2* = 1.6 Hz, 1H), 8.15-8.13 (m, 1H), 7.57-7.49 (m, 2H), 7.44-7.41 (m, 1H), 6.97-6.85 (m, 6H), 4.28 (s, 8H), 3.77 (dd, *J1* = 18 Hz, *J2* = 9.6 Hz, 2H), 3.31–3.36 (m, 1H), 2.76-2.69 (m, 1H), 2.38-2.31 (m, 1H); ¹³C NMR (100 MHz, CDCl3): *δ* 171.4, 148.0, 143.5, 142.3, 138.4, 136.5, 136.2, 134.5, 127.8, 127.4, 121.6, 121.5, 120.1, 117.3, 116.5, 115.8, 64.4, 64.4, 57.6, 39.0, 32.5; HRMS (ESI): calcd. for $C_{30}H_{27}N_2O_5 [M + H]^+$ 495.1920; found 495.1904.

2,4-bis(3,4-dichlorophenyl)-N-(quinolin-8-yl)cyclobutanecarboxamide (12j): Following the general procedure A described above, **12j** was obtained after purification by silica gel column chromatography ($EtOAc$: Hexane = 4:96) as white color solid (91 mg, 71%); R_f 0.40 (15% EtOAc/Hexane); mp 140-145^oC; IR (DCM): v_{max} 3341, 1631, 1587, 1332, 804 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.54 (s, 1H), 8.77 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.6$ Hz, 1H), 8.31 (dd, $J_1 = 7.6$ Hz, $J_2 = 1.2$ Hz, 1H), 8.13-8.10 (m, 1H), 7.45-7.41 (m, 4H), 7.36 (t, *J* = 8.0 Hz, 1H), 7.29-7.24 (m, 2H), 7.17- 7.14 (m, 2H), 4.12 (dd, *J*¹ = 8.0 Hz, *J*² = 3.2 Hz, 1H), 4.01-3.95 (m, 2H), 3.41 (dd, *J¹* $= 22.4$ Hz, $J_2 = 11.6$ Hz, 1H), 2.76-2.71 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 168.3, 148.1, 138.2, 136.4, 136.1, 134.0, 134.0, 132.6, 129.7, 128.8, 127.7, 127.0, 126.8, 121.5, 121.3, 116.2, 53.6, 37.4, 27.5; HRMS (ESI): *m/z* [M + H]⁺ calcd for $C_{26}H_{19}Cl_4N_2O$: 515.0251; found 515.0249.

2,4-bis(3,4-dimethylphenyl)-N-(quinolin-8-yl)cyclobutanecarboxamide (12k): Following the general procedure A described above, **12k** was obtained after purification by silica gel column chromatography (E tOAc : Hexane = 3 : 97) as pale yellow color solid (64 mg, 60%); R_f 0.40 (15% EtOAc/Hexane); mp 99-102 ^oC; IR (DCM): v_{max} 3435, 1575, 1365, 1291, 1047 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.47 $(s, 1H)$, 8.74 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.6$ Hz, 1H), 8.39-8.37 (m, 1H), 8.08-8.06 (m, 1H), 7.41-7.29 (m, 4H), 7.13-7.08 (m, 4H), 6.97 (d, *J* = 7.6 Hz, 2H), 4.11 (dd, *J*¹ = 8.4 Hz, $J_2 = 3.2$ Hz, 1H), 4.04-3.97 (m, 2H), 3.53-3.44 (m, 1H), 2.72-2.70 (m, 1H), 2.11 (s, 12H); ¹³C NMR (100 MHz, CDCl3): δ 169.4, 147.7, 138.3, 138.0, 136.1, 136.0, 134.4, 134.1, 129.3, 128.3, 127.7, 127.2, 124.4, 121.2, 120.7, 116.4, 54.7, 38.9, 30.3, 19.7, 19.3; HRMS (ESI): m/z [M + H]⁺ calcd for C₃₀H₃₁N₂O: 435.2423; found 435.2419.

N-(quinolin-8-yl)-2,4-di(thiophen-2-yl)cyclobutanecarboxamide (12l): Following the general procedure A described above, **12l** was obtained after purification by silica gel column chromatography (EtOAc : Hexane $= 5 : 95$) as yellow color solid (76 mg, 78%); R_f 0.30 (15% EtOAc/Hexane); mp 99-102 ^oC; IR (DCM): υ_{max} 3313, 1612, 1554, 1302, 801 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.60 (s, 1H), 8.75 (dd, $J_1 = 4.4$ Hz, *J*² = 1.6 Hz, 1H), 8.55-8.53 (m, 1H), 8.10-8.08 (m, 1H), 7.42-7.39 (m, 3H), 7.09- 7.05 (m, 4H), 6.89 (dd, *J*¹ = 4.8 Hz, *J*² = 3.6 Hz, 2H), 4.22-4.15 (m, 2H), 4.00-4,01 (m, 1H), 3.59-3.50 (m, 1H), 2.94-2.86 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 168.1, 147.9, 143.4, 138.2, 136.2, 134.3, 127.7, 127.3, 126.7, 125.2, 123.9, 121.4, 121.2, 116.6, 55.9, 35.7, 35.5; HRMS (ESI): m/z [M + H]⁺ calcd for C₂₂H₁₉N₂OS₂: 391.0938; found 391.0936.

2,6-diphenyl-N-(quinolin-8-yl)cyclohexanecarboxamide (12m): Following the general procedure A described above, **12m** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $4:96$) as white solid (38 mg, 33%);

R_f 0.5 (15% EtOAc/Hexane); mp 176-178 °C; IR (DCM): υ_{max} 3357, 2929, 2247, 1682, 1520, 1486, 1324, 1165 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.66 (s, 1H), 8.54 $(\text{dd}, J_1 = 7.6 \text{ Hz}, J_2 = 1.2 \text{ Hz}, 1\text{ H}), 8.45 \text{ (dd, } J_1 = 4.4 \text{ Hz}, J_2 = 1.6 \text{ Hz}, 1\text{ H}), 7.99 \text{ (dd, } J_1 = 1.6 \text{ Hz})$ = 8.4 Hz, *J²* = 1.6 Hz, 1H), 7.44–7.34 (m, 6H), 7.29–7.26 (m, 2H), 7.13 (t, *J* = 7.6 Hz, 4H), 6.98–6.94 (m, 2H), 3.22–3.18 (m, 3H), 2.87–2.77 (m, 2H), 2.30–2.24 (m, 1H), 1.86–1.82 (m, 2H), 1.70–1.67 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 171.0, 147.4, 144.1, 1378.0, 135.7, 134.1, 128.2, 127.5, 127.4, 127.0, 126.3, 121.1, 120.9, 115.9, 59.9, 47.9, 26.6, 25.6; HRMS (ESI): calcd. for $C_{28}H_{27}N_2O [M + H]^+$ 407.2123; found 407.2130.

2,6-bis(4-methoxyphenyl)-N-(quinolin-8-yl)cyclohexanecarboxamide (12n): Following the general procedure A described above, **12n** was obtained after purification by silica gel column chromatography ($EtOAc$: Hexane = 8 : 92) as pale yellow semi solid (34 mg, 30%); R*^f* 0.30 (15% EtOAc/Hexane); IR (DCM): υmax 3054, 2925, 2313, 1716, 1525, 1425, 1265, cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 8.65 (s, 1H), 8.52 (d, $J = 7.6$ Hz, 1H), 8.48 (dd, $J_I = 4.0$ Hz, $J₂ = 1.6$ Hz, 1H), 8.02-8.00 (m, 1H), 7.45–7.41 (m, 1H), 7.37-7.35 (m, 1H), 7.31-7.23 (m, 6H), 6.65 (d, *J* = 8.4 Hz, 4H), 3.54 (s, 6H), 3.16-3.10 (m, 3H), 2.74 (dd, *J¹* = 12.8 Hz, *J²* = 4.0 Hz, 2H), 2.25-2.21 (m, 1H), 1.82-1.78 (m, 2H), 1.68-1.65 (m, 1H); ¹³C NMR (100 MHz, CDCl3): *δ* 171.3, 157.9, 147.4, 138.0, 136.4, 135.7, 134.3, 128.4, 127.5, 127.0, 121.1, 120.8, 116.0, 113.6, 57.4, 55.0, 47.0, 26.7, 25.8; HRMS (ESI): calcd. for C₃₀H₃₁N₂O₃ $[M + Na]$ ⁺ 467.2335; found 467.2321.

2,3-bis(4-acetylphenyl)-N-(quinolin-8-yl)cyclopropanecarboxamide (12o): Following the general procedure A described above, **12o** was obtained after purification by silica gel column chromatography (EtOAc : Hexane $= 5 : 95$) as yellow solid (44 mg, 40%); R_f 0.40 (15% EtOAc/Hexane); mp 83 - 85 °C; IR (DCM): v_{max} 3346, 3053, 1682, 1525, 1486, 1325, 1162 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 10.00 (s, 1H), 8.72–8.70 (m, 1H), 8.61 (dd, $J_1 = 6.8$ Hz, $J_2 = 2.4$ Hz, 1H), 8.14 (dd, J_1 $= 8.4$ Hz, $J_2 = 1.6$ Hz, 1H), 7.80 (d, $J = 8.4$ Hz, 4H), 7.47–7.42 (m, 3H), 7.35 (d, $J =$ 8.4 Hz, 4H), 3.16 (d, *J* = 9.2 Hz, 2H), 2.91 (t, *J* = 9.2 Hz, 1H), 2.55 (s, 6H); ¹³C NMR (100 MHz, CDCl3): *δ* 198.0, 166.2, 148.0, 139.9, 138.1, 136.4, 135.3, 134.3, 131.2, 127.9, 127.6, 127.3, 121.6, 121.5, 116.5, 29.9, 29.4, 26.6; HRMS (ESI): calcd. for $C_{29}H_{25}N_2O_3$ [M + H]⁺ 449.1865; found 449.1848.

3-phenyl-N-(quinolin-8-yl)butanamide (14a): Following the general procedure A described above, **14a** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $4:96$) as light yellow liquid (57 mg, 78%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3353, 2959, 1680, 1510, 1480, 1322, 1162, 756 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.77 (s, 1H), 8.81-8.79 (m, 2H), 8.16 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H), 7.57–7.44 (m, 3H), 7.38-7.22 (m, 5H), 3.53 (dd, J_1 $= 14.8$ Hz, $J_2 = 6.8$ Hz, 1H), 2.95-2.90 (m, 1H), 2.80 (dd, $J_1 = 14.8$ Hz, $J_2 = 8.4$ Hz, 1H), 1.44 (d, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 170.4, 148.1, 146.0, 138.3, 136.3, 134.4, 128.6, 127.9, 127.4, 126.9, 126.4, 121.6, 121.5, 116.5, 46.9, 36.9, 21.9; HRMS (ESI): calcd. for $C_{19}H_{19}N_2O [M + H]^+$ 291.1497; found 291.1489.

3-(4-methoxyphenyl)-N-(quinolin-8-yl)butanamide (14b): Following the general procedure A described above, **14b** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 4 : 96) as pale yellow liquid (67 mg, 93%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3354, 2958, 1680, 1520, 1483, 1242, 1034, 826 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.73 (s, 1H), 8.79 (m, 2H), 8.16 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H), 7.56–7.44 (m, 4H), 7.27 (d, J = 8.4 Hz, 2H), 6.87 (d, J = 8.8 Hz, 2H), 3.78 (s, 3H), 3.50–3.45 (m, 1H), 2.90–2.74 (m, 2H), 1.41 (d, *J =* 6.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl3): *δ* 170.5, 158.1, 148.1, 138.3, 138.0, 136.3, 134.4, 127.9, 127.8, 127.4, 121.6, 121.4, 116.4, 114.0, 55.2, 47.2, 36.1, 22.1; HRMS (ESI): calcd. for $C_{20}H_{21}N_2O_2$ [M + H]⁺ 321.1603; found 321.1589.

N-(quinolin-8-yl)-3-(p-tolyl)butanamide (14c): Following the general procedure A described above, **14c** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $4:96$) as yellow semi solid (73 mg, 97%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3352, 2958, 1682, 1522, 1484, 1322, 1019, 824 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.77 (s, 1H), 8.81 (d, *J* = 6.8 Hz, 2H), 8.16 (d, *J* = 8.8 Hz, 1H), 7.57–7.44 (m, 3H), 7.29-7.25 (m, 2H), 7.15 (d, *J =* 7.6 Hz, 2H), 3.53-3.48 (m, 1H), 2.94-2.88 (m, 1H), 2.82-2.76 (m,1H), 2.33 (s, 3H), 1.43 (d, *J* $= 6.8$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 170.5, 148.1, 143.0, 138.3, 136.3, 135.9, 134.5, 129.3, 127.9, 127.4, 126.7, 121.6, 121.4, 116.5, 47.0, 36.5, 22.0, 21.0; HRMS (ESI): calcd. for $C_{20}H_{21}N_2O$ [M + H]⁺ 305.1654; found 305.1642.

3-(4-chlorophenyl)-N-(quinolin-8-yl)butanamide (14d): Following the general procedure A described above, **14d** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $4:96$) as pale yellow semi solid (61 mg, 75%); R*^f* 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3349, 2960, 1681, 1521, 1484, 1322, 1164, 1086, 825 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.72 (s, 1H), 8.79-8.75 (m, 2H),

8.15 (dd, *J¹* = 8.4 Hz, *J²* = 1.2 Hz, 1H), 7.53–7.44 (m, 3H), 7.28 (s, 4H), 3.53-3.48 (m, 1H), 2.89-2.76 (m, 2H), 1.41 (d, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃); δ 170.0, 148.1, 144.4, 138.2, 136.4, 134.3, 132.0, 128.7, 128.3, 127.9, 127.4, 121.6, 121.6, 116.5, 46.8, 36.3, 21.9; HRMS (ESI): calcd. for $C_{19}H_{18}CIN_2O$ $[M + H]^+$ 325.1105; found 325.1095.

N-(quinolin-8-yl)-3-(4-(trifluoromethoxy)phenyl)butanamide (14e): Following the general procedure A described above, **14e** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $4:96$) as colorless liquid (66 mg, 70%); R*^f* 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.73 (s, 1H), 8.79-8.76 (m, 2H), 8.16 (dd, *J¹* = 8.4 Hz, *J2* = 1.6 Hz, 1H), 7.54–7.44 (m, 3H), 7.37 (d, *J* = 8.8 Hz, 2H), 7.16 (d, *J* = 8.0 Hz, 2H), 3.55 (q, *J* = 7.2 Hz, 1H), 2.91-2.85 (m,1H), 2.83-2.77 (m, 1H), 1.43 (d, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 169.9, 148.1, 147.7, 147.7, 144.6, 138.2, 136.4, 134.3, 128.2, 127.9, 127.4, 121.6, 121.6, 121.1, 116.4, 46.8, 36.2, 21.8; HRMS (ESI): calcd. for $C_{20}H_{18}F_3N_2O_2$ [M + H]⁺ 375.1320; found 375.1304.

3-(4-acetylphenyl)-N-(quinolin-8-yl)butanamide (14f): Following the general procedure A described above, **14f** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 20 : 80) as brown semi solid (82 mg, 99%); R_f 0.10 (15% EtOAc/Hexane); IR (DCM): υmax 3349, 2964, 1682, 1525, 1486, 1325, 1164, 1014, 827 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.74 (s, 1H), 8.77-8.74 (m, 2H), 8.15 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H), 7.91 (d, $J = 8.4$ Hz, 2H), 7.5–7.43 (m, 5H), 3.62–3.56 (m, 1H), 2.92–2.80 (m, 2H), 2.56 (s, 3H), 1.44 (d, *J =* 7.2 Hz, 3H); ¹³C

NMR (100 MHz, CDCl₃): δ 197.9, 169.8, 151.6, 148.1, 138.2, 136.4, 135.5, 134.2, 128.8, 127.9, 127.4, 127.2, 121.6, 121.6, 116.5, 46.3, 36.9, 26.6, 21.7; HRMS (ESI): calcd. for $C_{21}H_{21}N_2O_2$ [M + H]⁺ 333.1603; found 333.1588.

3-(4-nitrophenyl)-N-(quinolin-8-yl)butanamide (14g): Following the general procedure A described above, **14g** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 4 : 96) as yellow solid (56 mg, 67%); R_f 0.40 (15% EtOAc/Hexane); mp 98-100 °C; IR (DCM): υ_{max} 3354, 2962, 1685, 1520, 1484, 1345, 1035, 845 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.73 (s, 1H), 8.77-8.75 (m, 1H), 8.72 (dd, $J_1 = 6.0$ Hz, $J_2 = 2.8$ Hz, 1H), 8.16 (d, $J = 8.8$ Hz, 3H), 7.52–7.44 (m, 5H), 3.65 (dd, *J¹* = 14.4 Hz, *J²* = 7.2 Hz, 1H), 2.88 (dd, *J¹* = 7.2 Hz, *J²* = 2.8 Hz, 2H), 1.45 (d, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 169.3, 153.6, 148.2, 146.6, 138.2, 136.4, 134.1, 127.9, 127.9, 127.3, 123.9, 121.7, 121.7, 116.5, 46.1, 36.8, 21.6; HRMS (ESI): calcd. for $C_{19}H_{18}N_3O_3$ [M + H]⁺ 336.1348; found 336.1335.

3-(4-cyanophenyl)-N-(quinolin-8-yl)butanamide (14h): Following the general procedure A described above, **14h** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 10 : 90) as brown semi solid (43 mg, 56%); R_f 0.10 (15% EtOAc/Hexane); IR (DCM): υmax 3345, 2965, 2229, 1680, 1521, 1485, 1163, 1016, 826 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.72 (s, 1H), 8.78 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.6$ Hz, 1H), 8.73 (dd, $J_1 = 6.0$ Hz, $J_2 = 2.8$ Hz, 1H), 8.16 (dd, $J_1 = 8.4$ Hz, J_2 = 1.6 Hz, 1H), 7.61–7.43 (m, 7H), 3.58 (q, *J =* 7.2 Hz, 1H), 2.85 (dd, *J¹* = 7.2 Hz, *J²* = 5.2 Hz, 2H), 1.43 (d, $J = 6.8$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 169.4, 151.5, 148.2, 138.2, 136.4, 134.1, 132.5, 127.9, 127.8, 127.4, 121.7, 119.0, 116.5, 110.3,

46.1, 36.9, 21.5; HRMS (ESI): calcd. for $C_{20}H_{18}N_3O [M + H]^+$ 316.1450; found 316.1440.

3-(4-fluorophenyl)-N-(quinolin-8-yl)butanamide (14i): Following the general procedure A described above, **14i** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 4 : 96) as colourless liquid (46 mg, 64%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3353, 2963, 1680, 1521, 1486, 1325, 1158, 1016, 826 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.72 (s, 1H), 8.79-8.76 (m, 2H), 8.18-815 (m, 1H), 7.56–7.45 (m, 3H), 7.33-7.29 (m, 2H), 7.00 (t, *J =* 8.8 Hz, 2H), 3.51 (q, *J* = 7.2 Hz, 1H), 2.86 (q, *J* = 7.2 Hz, 1H), 2.81-2.76 (m, 1H), 1.42 (d, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 170.1, 162.7, 160.2, 148.1, 141.5 (d, *J* = 12.0 Hz), 138.2, 136.4, 134.3, 128.3 (d, *J* = 31.6 Hz), 127.9, 127.4, 121.6 (d, *J* = 38.4 Hz), 116.4, 115.4 (d, $J = 83.0$ Hz), 47.0, 36.2, 22.0; HRMS (ESI): calcd. for C₁₉H₁₈ FN₂O $[M + H]$ ⁺ 309.1403; found 309.1388.

3-(3-nitrophenyl)-N-(quinolin-8-yl)butanamide (14j): Following the general procedure A described above, **14j** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $7:93$) as brown semi solid (72 mg, 86%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3348, 2965, 1682, 1531, 1486, 1350, 1164, 1031, 826 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.75 (s, 1H), 8.76 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.2$ Hz, 1H), 8.72 (dd, $J_1 = 6.4$ Hz, $J_2 = 2.8$ Hz, 1H), 8.23 (s, 1H), 8.14 (dd, J_1 = 8.4 Hz, *J²* = 1.6 Hz, 1H), 8.04 (dd, *J¹* = 8.0 Hz, *J²* = 1.6 Hz, 1H), 7.67 (d*, J* = 7.6 Hz, 1H), 7.51–7.43 (m, 4H), 3.68–3.63 (m, 1H), 2.95–2.84 (m, 2H), 1.47 (d*, J* = 6.8 Hz, 3H) ; ¹³C NMR (100 MHz, CDCl3): *δ* 169.4, 148.5, 148.2, 148.0, 138.2, 136.4, 134.1, 133.7, 129.5, 127.9, 127.3, 121.7, 121.6, 121.6, 116.5, 46.2, 36.5, 21.7 ; HRMS (ESI): calcd. for $C_{19}H_{18}N_3O_3$ [M + H]⁺ 336.1348; found 336.1333.

3-(3-bromophenyl)-N-(quinolin-8-yl)butanamide (14k): Following the general procedure A described above, **14k** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 4 : 96) as colorless liquid (77 mg, 84%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.74 (s, 1H), 8.80 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.6$ Hz, 1H), 8.77 (dd, $J_1 = 7.2$ Hz, $J_2 = 1.6$ Hz, 1H), 8.16 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H), 7.56–7.45 (m, 4H), 7.33 (d, *J =* 8.0 Hz, 1H), 7.27 (d, *J =* 9.2 Hz, 1H), 7.19-7.15 $(m, 1H)$, 3.52-3.47 $(m, 1H)$, 2.89 $(dd, J_1 = 14.8 \text{ Hz}, J_2 = 6.8 \text{ Hz}, 1H$), 2.78 $(dd, J_1 =$ 14.8 Hz, $J_2 = 8.0$ Hz, 1H), 1.42 (d, $J = 6.8$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 169.9, 148.3, 148.1, 138.3, 136.4, 134.3, 130.2 129.9, 129.6, 127.9, 127.4, 127.4, 125.8, 122.7, 121.6, 121.6, 116.5, 46.6, 36.6, 21.7; HRMS (ESI): calcd. for C₁₉H₁₈ $BrN_2O [M + Na]^+$ 369.0603; found 369.0590.

3-(3-chlorophenyl)-N-(quinolin-8-yl)butanamide (14l): Following the general procedure A described above, **14l** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 4 : 96) as pale yellow liquid (53 mg, 65%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.74 (s, 1H), 8.80 (dd, $J_1 = 4.0$ Hz, *J2* = 1.6 Hz, 1H), 8.78-8.76 (m, 1H), 8.16 (dd, *J¹* = 8.4 Hz, *J2* = 1.6 Hz, 1H), 7.56–7.45 (m, 3H), 7.35 (s, 1H), 7.24-7.18 (m, 2H), 3.51 (dd, *J¹* = 14.4 Hz, *J2* = 6.8 Hz, 1H), 2.89 (dd, $J_1 = 14.8$ Hz, $J_2 = 6.8$ Hz, 1H), 2.79 (dd, $J_1 = 14.8$ Hz, $J_2 = 8.0$ Hz, 1H), 1.42 (d, $J = 6.8$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 169.9, 148.1, 148.0, 138.2,

136.4, 134.4, 134.3, 129.9, 127.9, 127.4, 127.0, 126.6, 125.3, 121.6, 121.6, 116.5, 46.6, 36.6, 21.7; HRMS (ESI): calcd. for $C_{19}H_{18}CIN_2O$ $[M + H]^+$ 325.1108; found 325.1093.

N-(quinolin-8-yl)-3-(m-tolyl)butanamide (14m): Following the general procedure A described above, **14m** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 4 : 96) as colorless liquid (63 mg, 83%); R_f 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.77 (s, 1H), 8.82-8.79 (m, 2H), 8.16 (dd, *J¹* = 8.0 Hz, *J²* = 1.6 Hz, 1H), 7.57–7.44 (m, 3H), 7.25-7.16 (m, 3H), 7.04 (d, *J =* 7.2 Hz, 1H), 3.53-3.47 (m, 1H), 2.95-2.90 (m, 1H), 2.79 (dd, $J_1 = 14.4$ Hz, $J_2 = 8.4$ Hz, 1H), 2.36 (s, 3H), 1.44 (d, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 170.5, 148.1, 146.0, 138.3, 138.2, 136.3, 134.4, 128.5, 127.9, 127.7, 127.4, 127.2, 123.9, 121.6, 121.4, 116.4, 46.9, 36.9, 21.9, 21.5; HRMS (ESI): calcd. for $C_{20}H_{21}N_{2}O$ [M + H]⁺ 305.1654; found 305.1640.

3-(3,4-dichlorophenyl)-N-(quinolin-8-yl)butanamide (14n): Following the general procedure A described above, **14n** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $5: 95$) as pale yellow solid (45 mg, 50%); R_f 0.40 (15% EtOAc/Hexane); mp 85-87 °C; IR (DCM); v_{max} 3412, 2965, 1684, 1526, 1485, 1325, 1122, 1039, 825 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.70 (s, 1H), 8.79 (dd, $J_1 = 4.0$ Hz, $J_2 = 1.6$ Hz, 1H), 8.74 (dd, $J_1 = 6.8$ Hz, $J_2 = 2$ Hz, 1H), 8.16 (dd, $J_1 =$ 8.4 Hz, *J²* = 1.6 Hz, 1H), 7.56–7.44 (m, 4H), 7.35 (d, *J* = 8.4 Hz, 1H), 7.17 (dd, *J¹* = 8.4 Hz, *J²* = 2.4 Hz, 1H), 3.51–3.46 (m, 1H), 2.88–2.76 (m, 2H), 1.41 (d*, J* = 6.8 Hz, 3H) ; ¹³C NMR (100 MHz, CDCl3): *δ* 169.6, 148.2, 146.2, 138.2, 136.4, 134.2, 132.5, 130.5, 130.3, 128.9, 127.9, 127.3, 126.6, 121.6, 116.5, 46.5, 36.2, 21.7; HRMS (ESI): calcd. for $C_{19}H_{17}Cl_2N_2O$ $[M + H]^+$ 359.0718; found 359.0706.

3-(4-bromo-3-fluorophenyl)-N-(quinolin-8-yl)butanamide (14o): Following the general procedure A described above, **14o** was obtained after purification by silica gel column chromatography (EtOAc : Hexane $= 4$: 96) as pale yellow solid (71 mg, 74%); R_f 0.40 (15% EtOAc/Hexane); mp 146-148 °C; IR (DCM): υ_{max} 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.71 (s, 1H), 8.79 (dd, *J¹* = 4.4 Hz, *J2* = 1.6 Hz, 1H), 8.75-8.73 (m, 1H), 8.18-815 (m, 1H), 7.56–7.43 (m, 4H), 7.13 (dd, *J¹* = 9.6 Hz, *J2* = 2.0 Hz, 1H), 7.02 (dd, *J¹* = 8.0 Hz, *J2* = 2.0 Hz, 1H), 3.50 (q, *J* = 7.2 Hz, 1H), 2.88-2.83 (m, 1H), 2.79 (dd, *J¹* = 14.8 Hz, *J2* = 7.6 Hz, 1H), 1.41 (d, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 169.6, 160.4, 157.9, 148.2, 147.8 (d, *J* = 25.2 Hz), 138.2, 136.4, 134.2, 133.5, 127.9, 127.4, 124.0 (d, $J = 12.4$ Hz), 121.7, 116.5, 115.0 (d, $J = 87.6$ Hz), 106.5 (d, $J = 83.6$ Hz), 46.4, 36.3, 36.3, 21.7; HRMS (ESI): calcd. for C₁₉H₁₇BrFN₂O [M + H]⁺ 387.0508; found 387.0495.

3-(4-methoxyphenyl)-N-(quinolin-8-yl)hexanamide (14p): Following the general procedure A described above, **14p** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $7 : 93$) as red solid (92 mg, 92%); R_f 0.40 (15%) EtOAc/Hexane); mp 85-87 °C; IR (DCM): υ_{max} 3355, 2956, 1683, 1525, 1485, 1325, 1248, 1035 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.69 (s, 1H), 8.77-8.75 (m, 2H), 8.13 (d, *J* = 8.0 Hz, 1H), 7.54–7.42 (m, 3H), 7.23 (d*, J* = 8.4 Hz, 2H), 6.85 (d*, J* = 8.4 Hz, 2H), 3.75 (s, 3H), 3.31–3.27 (m, 1H), 2.85–2.82 (m, 2H), 1.79–1.66 (m, 2H), 1.29–

1.22 (m, 2H), 0.89 (t, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 158.1, 148.0, 138.3, 136.3, 136.3, 134.5, 128.5, 127.9, 127.4, 121.5, 121.4, 116.4, 113.9, 55.2, 46.2, 41.6, 38.6, 20.6, 14.0; HRMS (ESI): calcd. for $C_{22}H_{25}N_2O_2$ [M + H]⁺ 349.1916; found 349.1906.

N-(quinolin-8-yl)-3-(p-tolyl)hexanamide (14q): Following the general procedure A described above, **14q** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $5:95$) as pale yellow semi solid (77 mg, 93%); R*^f* 0.40 (15% EtOAc/Hexane); IR (DCM): υmax 3356, 2956, 1688, 1525, 1485, 1325, 1160, 1114, 825 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.72 (s, 1H), 8.79–8.77 (m, 2H), 8.14 (dd, *J¹* = 8.4 Hz, *J2* = 1.6 Hz, 1H), 7.54–7.43 (m, 3H), 7.21 (d*, J* = 8.0 Hz, 2H), 7.13 (d*, J* = 8.0 Hz, 2H), 3.33–3.30 (m, 1H), 2.86 (d*, J* = 6.8 Hz, 2H), 2.31 (s, 3H), 1.80–1.68 (m, 2H), 1.29–1.22 (m, 2H), 0.89 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl3): *δ* 170.6, 148.0, 141.3, 138.3, 136.3, 135.8, 134.5, 129.2, 127.9, 127.4, 127.4, 121.5, 121.3, 116.4, 46.0, 42.0, 38.5, 21.1, 20.6, 14.1; HRMS (ESI): calcd. for $C_{22}H_{25}N_2O$ [M + H]⁺ 333.1967; found 333.1960.

3-(4-methoxyphenyl)-N-(quinolin-8-yl)heptanamide (14r): Following the general procedure A described above, **14r** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $5: 95$) as pale yellow solid (94 mg, 97%); R_f 0.40 (15% EtOAc/Hexane); mp 89-91 °C; IR (DCM): v_{max} 3356, 2929, 1682, 1525, 1486, 1325, 1247, 1036 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.68 (s, 1H), 8.77–8.74

(m, 2H), 8.15 (d, *J* = 8.0 Hz, 1H), 7.54–7.43 (m, 3H), 7.23 (d, *J* = 8.4 Hz, 2H), 6.85 (d*, J* = 8.8 Hz, 2H), 3.76 (s, 3H), 3.28–3.24 (m, 1H), 2.85–2.81 (m, 2H), 1.80–1.67 (m, 3H), 1.32–1.17 (m, 5H), 0.90–0.83 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 158.0, 148.0, 138.3, 136.4, 136.3, 134.4, 128.4, 127.8, 127.4, 121.5, 121.3, 116.4, 113.9, 55.2, 46.2, 41.8, 36.1, 29.7, 22.7, 14.0; HRMS (ESI): calcd. for $C_{23}H_{27}N_2O_2$ [M + H]⁺ 363.2073; found 363.2082.

3-(4-methoxyphenyl)-N-(quinolin-8-yl)nonanamide (14s): Following the general procedure A described above, **14s** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $7:93$) as pale yellow solid (93 mg, 95%); R_f 0.40 (15% EtOAc/Hexane); mp 75-77 °C; IR (DCM): υ_{max} 3356, 2928, 1683, 1520, 1486, 1325, 1247, 1036 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.68 (s, 1H), 8.78–8.75 (m, 2H), 8.14 (d, *J* = 8.4 Hz, 1H), 7.54–7.43 (m, 3H), 7.23 (d, *J* = 8.8 Hz, 2H), 6.85 (d*, J* = 8.8 Hz, 2H), 3.76 (s, 3H), 3.28–3.25 (m, 1H), 2.85–2.81 (m, 2H), 1.79–1.67 (m, 2H), $1.28-1.22$ (m, 8H), 0.85 (t, $J = 6.8$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 158.0, 148.0, 138.3, 136.4, 136.3, 134.5, 128.4, 127.9, 127.4, 121.5, 121.3, 116.4, 113.9, 55.2, 46.2, 41.9, 36.4, 31.8, 29.3, 27.4, 22.6, 14.1; HRMS (ESI): calcd. for $C_{25}H_{31}N_2O_2$ [M + H]⁺ 391.2386; found 391.2379.

3-(4-methoxyphenyl)-N-(quinolin-8-yl)dodecanamide (14t): Following the general procedure A described above, **14t** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $7:93$) as pale yellow solid (91 mg, 85%); R_f 0.40 (15% EtOAc/Hexane); mp 56-58 °C; IR (DCM): υ_{max} 3356, 2926, 1683, 1520, 1486, 1325, 1248, 1037 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.68 (s, 1H), 8.77–8.75 (m, 2H), 8.13 (d, *J* = 8.4 Hz, 1H), 7.51–7.43 (m, 3H), 7.23 (d, *J* = 8.8 Hz, 2H), 6.85 (d*, J* = 8.8 Hz, 2H), 3.76 (s, 3H), 3.30–3.25 (m, 1H), 2.85–2.81 (m, 2H), 1.80–1.67 (m, 2H), 1.30–1.22 (m, 15H), 0.89 (t, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): *δ* 170.6, 158.0, 148.0, 138.3, 136.4, 136.3, 134.5, 128.4, 127.9, 127.4, 121.5, 121.3, 116.4, 113.9, 55.1, 46.2, 41.9, 36.4, 31.9, 29.6, 29.6, 29.6, 29.3, 27.5, 22.7, 14.2; HRMS (ESI): calcd. for $C_{28}H_{37}N_2O_2$ [M + H]⁺ 433.2855; found 433.2838.

3-(4-methoxyphenyl)-3-phenyl-N-(quinolin-8-yl)propanamide (14u): Following the general procedure A described above, **14u** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $8:92$) as brown semi solid (76 mg, 80%); R*^f* 0.20 (15% EtOAc/Hexane); IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.80 (s, 1H), 8.78-8.75 (m, 2H), 8.12 (dd, *J¹* = 8.4 Hz, *J²* = 1.6 Hz, 1H), 7.51–7.29 (m, 9H), 7.21 (t, *J =* 7.2 Hz, 1H), 6.86 (d, *J* = 8.4 Hz, 2H), 4.79 (t, *J =* 7.6 Hz, 1H), 3.75 (s, 3H), 3.33 (d, *J* = 7.6 Hz, 2H); ¹³C NMR (100 MHz, CDCl3): *δ* 169.7, 158.2, 148.1, 144.2, 138.2, 136.3, 135.9, 134.4, 128.8, 128.7, 127.9, 127.7, 127.4, 126.5, 121.6, 121.5, 116.5, 114.0, 55.2, 46.4, 44.7; HRMS (ESI): calcd. for $C_{25}H_{23}N_2O_2$ [M + Na]⁺ 383.1760; found 383.1745.

3-(4-methoxyphenyl)-N-(quinolin-8-yl)propanamide (14v): Following the general procedure A described above, **14v** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 4 : 96) as brown semi solid (13 mg, 17%); R_f 0.20 (15% EtOAc/Hexane); IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.80 (s, 1H), 8.82-8.79 (m, 2H), 8.18 (dd, $J_1 = 8.4$ Hz, $J_2 = 1.6$ Hz, 1H), 7.56–7.45 (m, 3H), 7.29-7.23 (m, 2H), 6.86 (d, $J =$ 8.8 Hz, 2H), 3.80 (s, 3H), 3.14-3.10 (m, 2H), 2.90-2.86 (m, 2H); ¹³C NMR (100 MHz, CDCl3): *δ* 170.9, 158.0, 148.1, 138.3, 136.3, 134.4, 132.8, 129.4, 127.9, 127.4, 121.6, 121.5, 116.5, 114.0, 55.3, 40.1, 30.7; HRMS (ESI): calcd. for $C_{19}H_{19}N_2O_2$ [M + H]⁺ 307.1447; found 307.1436.

3,3-bis(4-methoxyphenyl)-N-(quinolin-8-yl)propanamide (14v'): Following the general procedure A described above, **14v'** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $8:92$) as brown semi solid (60 mg, 58%); R*^f* 0.20 (15% EtOAc/Hexane; IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.76 (s, 1H), 8.78-8.76 (m, 1H), 8.73 (dd, *J¹* = 6.8 Hz, *J²* = 2.0 Hz, 1H), 8.16-8.14 (m, 1H), 7.51–7.43 (m, 3H), 7.29-7.25 (m, 4H), 6.84 (d, *J* = 8.4 Hz, 4H), 4.68-4.72 (m, 1H), 3.75 (s, 6H), 3.27 (d, $J = 7.6$ Hz, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 169.8, 158.1, 148.0, 138.2, 136.3, 136.3, 134.4, 128.7, 127.8, 127.4, 121.6, 121.4, 116.5, 114.0, 55.2, 45.6, 44.9; HRMS (ESI): calcd. for $C_{26}H_{25}N_2O_3$ [M + H]⁺ 413.1865; found 413.1848.

2,6-diheptyl-N-(quinolin-8-yl)benzamide (12t): Following the general procedure A described above, **12t** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $2:98$) as brown semi solid (62 mg, 45%); R_f 0.8 (15% EtOAc/Hexane); IR (DCM υmax 3348, 2925, 2855, 1677, 1522, 1480, 1325, 1262, 1113 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.96 (s, 1H), 9.04-9.01 (m, 1H), 8.76-8.74 (m, 1H), 8.21 (dd, *J¹* = 8.0 Hz, *J2* = 1.6 Hz, 1H), 7.66-7.58 (m, 2H), 7.46 (dd, $J_1 = 8.4$ Hz, $J_2 = 4.4$ Hz, 1H) 7.36-7.32 (m, 1H), 7.18 (d, $J = 8.0$ Hz, 2H), 2.73 (t, *J =* 8.0 Hz, 4H), 1.73-1.65 (m, 4H), 1.30-1.12 (m, 18H), 0.79-0.75 (m, 6H); ¹³C NMR (100 MHz, CDCl3): *δ* 168.9, 148.2, 139.5, 138.5, 137.5, 136.3, 134.5, 129.0, 128.0, 127.5, 126.7, 121.8, 121.8, 121.6, 116.7, 33.5, 31.7, 31.6, 29.6, 29.0, 22.6, 14.0; HRMS (ESI): calcd. for $C_{30}H_{41}N_2O$ [M + H]⁺ 445.3219; found 445.3236.

2,6-dibutyl-N-(quinolin-8-yl)benzamide (12u): Following the general procedure A described above, **12u** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = $2:98$) as brown semi solid (49 mg, 55%); R_f 0.8 (15% EtOAc/Hexane); IR (DCM): υmax 3347, 2927, 2858, 1676, 1522, 1482, 1325, 1262, 1126, cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 9.95 (s, 1H), 9.01 (dd, *J₁* = 7.6 Hz, *J2* = 1.2 Hz, 1H), 8.75 (dd, *J¹* = 4.4 Hz, *J2* = 1.6 Hz, 1H), 8.22-8.20 (m, 1H), 7.67- 7.58 (m, 2H), 7.48-7.45 (m, 1H), 7.36-7.32 (m, 1H), 7.18 (d, *J* = 7.6 Hz, 2H), 2.73 (t, *J =* 8.0 Hz, 4H), 1.73-1.65 (m, 5H), 1.32 (dd, *J¹* = 14.8 Hz, *J2* = 7.6 Hz, 4H), 0.84- 0.80 (m, 6H); ¹³C NMR (100 MHz, CDCl3): *δ* 168.9, 148.2, 139.5, 138.5, 137.5, 136.3, 134.5, 128.9, 128.0, 127.5, 126.7, 121.9, 121.7, 116.8, 33.8, 33.2, 22.7, 13.9; HRMS (ESI): calcd. for $C_{24}H_{29}N_2O$ [M + H]⁺ 316.2280; found 316.2265.

methyl 3-phenylbutanoate (15a): Following the general procedure B described above, **15a** was obtained after purification by silica gel column chromatography (EtOAc : Hexane = 1 : 99) as pale yellow liquid $(31 \text{ mg}, 37\%)$; R_f 0.80 (5%) EtOAc/Hexane); IR (DCM): υmax 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.35-7.22 (m, 5H), 3.65 (s, 3H), 3.32 (dd, J_1 = 14.8 Hz, *J²* = 7.2 Hz, 1H), 2.66 (dd, *J¹* = 15.2 Hz, *J²* = 6.8 Hz, 1H), 2.57 (dd, *J¹* = 15.2 Hz, $J_2 = 8.4$ Hz, 1H, 1.33 (d, $J = 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 172.9, 145.7, 128.5, 126.7, 126.4, 51.5, 42.7, 36.4, 21.8; HRMS (ESI): calcd. for $C_{11}H_{14}NaO_2$ [M + Na]⁺ 201.0891; found 201.0883.

3-phenylbutanoic acid (15b): Following the general procedure C described above, **15b** was obtained (EtOAc : Hexane = $1:99$) as brown semi solid (16 mg, 32%); IR (DCM): υ_{max} 3339, 2923, 2247, 1716, 1510, 1466, 1242, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl3): *δ* 7.35-7.22 (m, 5H), 3.30 (dd, *J¹* = 14.8 Hz, *J²* = 7.2 Hz, 1H), 2.70 (dd, $J_1 = 15.2$ Hz, $J_2 = 6.8$ Hz, 1H), 2.61 (dd, $J_1 = 15.6$ Hz, $J_2 = 8.4$ Hz, 1H), 1.35 (d, *J* $= 7.2$ Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 178.1, 145.5, 128.6, 126.7, 126.5, 42.5, 36.2, 21.9; HRMS (ESI): calcd. for $C_{10}H_{11}O_2$ [M - H]⁺ 163.0759; found 163.0752.

2,4-Bis(4-bromophenyl)cyclobutanecarboxylic acid (15c). Following the general procedure C described above, **15c** was obtained as a brown color solid (crude material was almost pure); 65 mg, 52% yield; mp131-133 °C; IR (DCM): v_{max} 2922, 1698,

1516, 1425, 806 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.48 (d, *J* = 8.4 Hz, 4H), 7.20 $(d, J = 8.4 \text{ Hz}, 4\text{H})$, 3.76 (dd, $J_1 = 18.4 \text{ Hz}, J_2 = 9.6 \text{ Hz}, 2\text{H})$, 3.24 (t, $J = 9.6 \text{ Hz}, 1\text{H}$), 2.80 (dd, $J_1 = 18.8$ Hz, $J_2 = 8.4$ Hz, 1H), 2.26 (dd, $J_1 = 20.8$ Hz, $J_2 = 10.4$ Hz); ¹³C NMR (100 MHz, CDCl3): δ 179.4, 141.4, 131.6, 128.4, 120.6, 52.2, 38.8, 32.4; HRMS (ESI): m/z [M + Na]⁺ calcd for C₁₇H₁₄Br₂O₂Na: 430.9258; found 430.9281.

2,4-Bis(4-chlorophenyl)cyclobutanecarboxylic acid (15d). Following the general procedure C described above, **15d** was obtained as a white colour liquid (crude material was almost pure); 83 mg, 59% yield; IR (DCM): υmax 2966, 1565, 1432, 1331, 800 cm⁻¹; ¹H NMR (400 MHz, CDCl₃): δ 7.35 (d, *J* = 8.4 Hz, 4H), 7.25 (d, *J* = 8.4 Hz, 4H), 3.80 (dd, *J*¹ = 18.4 Hz, *J*² = 9.6 Hz, 2H), 3.27 (t, *J* = 9.6 Hz, 1H), 2.82 (dd, $J_1 = 18.8$ Hz, $J_2 = 8.4$ Hz, 1H), 2.28 (dd, $J_1 = 21.2$ Hz, $J_2 = 10.8$ Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 179.7, 140.9, 132.6, 128.7, 128.0, 52.4, 38.8, 32.5; FT-IR (DCM): 2966, 1565, 1432, 1331, 800 cm⁻¹; HRMS (ESI): m/z [M - H] calcd for $C_{17}H_{13}Cl_2O_2$: 319.0292; found 319.0298.

Summary

In summary, Pd-catalyzed Ag_2CO_3 mediated one-pot multicomponent reaction method for DG installation, C-H activation and β- arylation of $sp³$ C-H bonds of various carboxamide derivatives were successfully achieved. The multicomponent reaction of a mixture of a bidentate ligand, acid chloride and aryl iodide in the presence of Pd(OAc)₂ catalyst and Ag₂CO₃ additive directly afforded the corresponding β-arylated N-(quinolin-8-yl)carboxamide derivative. The bidentate ligand 8-aminoquinoline was found to be the best DG to promote the one-pot Pd-catalyzed C-H activation reaction on $sp³$ C-H bonds. Various types of acid chlorides and aryl iodides were used to check the efficiency of the reaction. We have explored a step economical and straightforward synthetic methodology via one-pot manner for the synthesis of a variety of β-arylated carboxamide derivatives in moderate to high yields.

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Appendix NMR Spectra of Representative Compounds

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SpinWorks 4: SM 182 B2
PROTON CDCI3 /opt/topspin3.5pl2/nmrdata nmrsu 4 ∰ס (ס פס פס פס פס פס)
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10000771885
1000771885
1000771884

SpinWorks 4: SM 203 A2
PROTON CDCl3 /opt/topspin3.5pl2/nmrdata nmrsu 1

22222323
52231147
22231247

SpinWorks 4: SM 206 B2
PROTON CDCl3 /opt/topspin3.5pl2/nmrdata nmrsu 4

SpinWorks 4: SM-214-A2

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 $\begin{array}{c}\n\hline\n1 \\
80\n\end{array}$

 $\begin{array}{c}\n\hline\n\end{array}$

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40\n\end{array}$

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 $\begin{array}{c}\n\hline\n1 \\
160\n\end{array}$

PPM

 $\begin{array}{c}\n\hline\n1 \\
140\n\end{array}$

 $\begin{array}{c}\n1 \\
120\n\end{array}$

 $\begin{array}{c}\n\hline\n1 \\
100\n\end{array}$

SpinWorks 4: SM 275 A2
PROTON CDCl3 /opt/topspin3.5pl2/nmrdata nmrsu 21

SpinWorks 4: sm 160 b2

SpinWorks 4: sm 257 a2
PROTON CDCl3 /opt/topspin3.5pl2/nmrdata nmrsu 53

Br,

 $\begin{array}{c}\n 3.722 \\
 3.745 \\
 3.767\n \end{array}$ $\frac{3.214}{3.262}$ $\begin{array}{c}\n 2.791 \\
 2.817 \\
 2.819\n\end{array}$ $\begin{array}{c}\n 2.218 \\
 2.2744 \\
 2.2796 \\
 2.296\n \end{array}$

SpinWorks 4: SM 272 A
PROTON CDCl3 /opt/topspin3.5pl2/nmrdata nmrsu 8

 $\begin{array}{c}\n\hline\n1 \\
120\n\end{array}$

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 $\begin{array}{c}\n\hline\n1 \\
160\n\end{array}$

 PPM

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100\n\end{array}$

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